Early signals of significantly increased vaccine breakthrough, decreased hospitalization rates, and less severe disease in patients with COVID-19 caused by the Omicron variant of SARS-CoV-2 in Houston, Texas

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Abstract

Genetic variants of SARS-CoV-2 continue to dramatically alter the landscape of the COVID-19 pandemic. The recently described variant of concern designated Omicron (B.1.1.529) has rapidly spread worldwide and is now responsible for the majority of COVID-19 cases in many countries. Because Omicron was recognized very recently, many knowledge gaps exist about its epidemiology and clinical severity and disease course. A comprehensive genome sequencing study of SARS-CoV-2 in the Houston Methodist healthcare system identified 862 symptomatic patients with infections caused by Omicron from late November 2021 through December 18, 2021. Omicron very rapidly increased in only three weeks to cause 90% of all new COVID-19 cases. Compared to patients infected with either Alpha or Delta variants in our healthcare system, Omicron patients were significantly younger, had significantly increased vaccine breakthrough rates, and were significantly less likely to be hospitalized. Omicron patients required less intense respiratory support and had a shorter length of hospital stay, consistent with decreased disease severity. Although the number of Omicron patients we studied is relatively small, in the aggregate the data document the unusually rapid spread and increased occurrence of COVID-19 caused by the Omicron variant in metropolitan Houston, and provide information about disease character.

[Introduction]

Over the last 14 months, the Alpha and Delta variants of concern (VOCs) of SARS-CoV-2 have caused two distinct COVID-19 disease surges in the United States, Southeast Asia, Europe, and elsewhere (https://www.cdc.gov/coronavirus/2019-ncov/cases-updates/variant-surveillance/variant-info.html, last accessed December 30, 2021; https://www.gov.uk/government/collections/new-sars-cov-2-variant, last accessed December 30, 2021)[1], and remodeled the landscape of human behavior and many societies. Delta replaced the Alpha variant as the cause of virtually all COVID-19 in many countries (https://www.who.int/publications/m/item/weekly-epidemiological-update-on-covid-19---13-july-2021, last accessed August 18, 2021; https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/conditionsanddiseases/bulletins/coronaviruscovid19infectionsurveypilot/9july2021, last accessed August 18, 2021).

At the start of the pandemic almost two years ago, the Houston Methodist healthcare system instituted a comprehensive and integrated population genomics project designed to sequence all SARS-CoV-2 samples causing COVID-19 in patients cared for at our facilities, which include eight hospitals located throughout the metroplex. The project was implemented when the initial Houston Methodist COVID-19 case was diagnosed at the end of February 2020, and has continued unabated[2-6]. This project was facilitated by the existence of a single large diagnostic laboratory that serves the entire system and is seamlessly integrated with a research institute with extensive genomics expertise and capacity. A key goal was to comprehensively map the population genomics, trajectory, and other features of the pandemic in metropolitan

Houston with a population size of approximately 7.2 million. Houston is the fourth largest city in the United States, the most ethnically diverse metropolitan area in the country, and is a major port of entry. To date, SARS-CoV-2 genomes have been sequenced from greater than 70,000 patient samples. Many features of four distinct SARS-CoV-2 waves in Houston have been described[2-6].

The successes of rapid SARS-CoV-2 vaccine development and documented efficacy, coupled with the significant downturn of the disease wave caused by Delta in Houston and elsewhere in fall, 2021[6], suggested that the pandemic was abating. However, the identification of a new VOC designated B.1.1.529 and commonly known as Omicron that has spread rapidly in South Africa and the UK has tempered this optimism[7-9]. Inasmuch as Omicron was recognized very recently, and much is not known about its epidemiology and clinical characteristics and course, we used our integrated infrastructure in an effort to address some pertinent knowledge gaps. Genome sequencing identified 862 COVID-19 patients with disease caused by Omicron in the Houston Methodist healthcare system beginning in late November 2021 and ending December 18, 2021. In three weeks Omicron spread throughout the Houston metropolitan region and became the cause of 90% of new COVID-19 cases. Although the number of our Omicron patients is relatively small, compared to patients infected with either Alpha or Delta variants and cared for in our system, significantly fewer Omicron patients were hospitalized, and those who were hospitalized required significantly less intense respiratory support and had a shorter length of stay. We cautiously interpret our findings to be consistent with decreased disease severity among Houston Methodist Omicron patients. Many factors undoubtedly have contributed,

including but not limited to increased vaccination uptake, population immunity, and patient demographics such as younger age. The extent to which our findings translate to other cities and other patient populations, including children, is unknown. Our study was not designed to address possible intrinsic differences in virulence of Omicron compared to Alpha, Delta, and other VOCs. We believe this topic remains an open question that warrants further investigation.

Materials and Methods

Patient Specimens

Specimens were obtained from patients registered at Houston Methodist facilities (e.g., hospitals and urgent care centers), and institutions in the Houston metropolitan region that use our laboratory services. The great majority of individuals had signs or symptoms consistent with COVID-19 disease. For analyses focusing on patients with COVID-19 caused by the Omicron variant, samples obtained from November 27, 2021 through December 18, 2021 were used. This time frame was chosen because it represents the period during which an Omicron variant was first identified in our healthcare system and the last date of specimen collection used to generate genome sequence data for this manuscript. For analyses comparing features of patients infected with the Omicron VOC and Alpha and Delta VOCs, all patients documented to be

131 infected with these variants in the Houston Methodist system were studied. The study
132 included 36,164 unique patients identified in this time frame for whom we had SARS-
133 CoV-2 genome sequences. The work was approved by the Houston Methodist
134 Research Institute Institutional Review Board (IRB1010-0199).
135
136 ## SARS-CoV-2 Molecular Diagnostic Testing
137
138 Specimens obtained from symptomatic patients with a suspicion for COVID-19 disease
139 were tested in the Molecular Diagnostics Laboratory at Houston Methodist Hospital
140 using assays granted Emergency Use Authorization (EUA) from the FDA
141 (https://www.fda.gov/medical-devices/emergency-situations-medical-devices/faqs-
142 diagnostic-testing-sars-cov-2#offeringtests, last accessed June 7, 2021). Multiple
143 molecular testing platforms were used, including the COVID-19 test or RP2.1 test with
144 BioFire Film Array instruments, the Xpert Xpress SARS-CoV-2 test using Cepheid
145 GeneXpert Infinity or Cepheid GeneXpert Xpress IV instruments, the Cobas SARS-
146 CoV-2 & Influenza A/B Assay using the Roche Liat system, the SARS-CoV-2 Assay
147 using the Hologic Panther instrument, the Aptima SARS-CoV-2 Assay using the Hologic
148 Panther Fusion system, the Cobas SARS-CoV-2 test using the Roche 6800 system,
149 and the SARS-CoV-2 assay using Abbott Alinity m instruments. Virtually all tests were
150 performed on material obtained from nasopharyngeal swabs immersed in universal
151 transport media (UTM); oropharyngeal or nasal swabs, bronchoalveolar lavage fluid, or
152 sputum treated with dithiothreitol (DTT) were sometimes used. Standardized specimen

collection methods were used (https://vimeo.com/396996468/2228335d56, last accessed June 7, 2021).

SARS-CoV-2 Genome Sequencing, Genome Analysis, and Identification of Variants

We sequenced the SARS-CoV-2 genome of >90% of all positive cases in the Houston Methodist healthcare system during the period studied. Libraries for whole SARS-CoV-2 genome sequencing were prepared according to version 4 (https://community.artic.network/t/sars-cov-2-version-4-scheme-release/312, last accessed August 19, 2021) of the ARTIC nCoV-2019 sequencing protocol. The semi-automated workflow used has been described previously[2-6]. Sequence reads were generated with an Illumina NovaSeq 6000 instrument.

Viral genomes were assembled with the BV-BRC SARS-Cov2 assembly service (https://www.bv-brc.org/app/ComprehensiveSARS2Analysis, last accessed June 7, 2021, requires registration). The pipeline currently uses seqtk version 1.3-r117 for sequence trimming (https://github.com/lh3/seqtk.git) and minimap version 2.17 for aligning reads against the Wuhan-Hu-1 (NC_045512.2) reference genome. Samtools version 1.11 was used for sequence and file manipulation, where maximum depth and minimum depth parameters in mpileup were set to 8,000 and 3, respectively. iVar version 1.3.1 was used for primer trimming and variant calling. Genetic lineages, VOCs, and variants of interest (VOIs) were identified based on genome sequence data and designated by Pangolin v. 3.1.17 with pangoLEARN module 2021-12-06

(https://cov-lineages.org/resources/pangolin.html, last accessed December 12, 2021). Genome data used in this study have been deposited to GISAID www.gisaid.org.

S-Gene Target-Failure Assay

An S-gene target-failure (SGTF) assay (TaqPath COVID-19 Combo Kit Thermo Fisher, Inc.), was used as a surrogate marker for the Omicron VOC for some specimens collected between December 18, 2021 and December 22, 2021. From November 1, 2021 onward, only Delta and Omicron were circulating in metropolitan Houston, as documented by whole-genome sequence data. Thus, samples with S-gene target failure were classified as Omicron, whereas samples yielding amplification of the S-gene were classified as a Delta variant.

Patient Metadata and Geospatial Analysis

Patient metadata were acquired from the electronic medical record by standard informatics methods. Figures showing geospatial distribution of spread for Omicron were generated with Tableau version 2021.2.7 (Tableau Software, LLC, Seattle, WA) using patient home address zip codes. A vaccination breakthrough case was defined as a PCR-positive sample from a patient obtained greater than 14 days after full vaccination (e.g., both doses of the Pfizer or Moderna mRNA vaccines) was completed. A booster vaccination breakthrough case was defined as a PCR-positive sample from a patient obtained greater than 14 days after receiving a third vaccine dose. For some

cases, manual chart review was conducted to resolve discrepancies or clarify ambiguities.

Results

Omicron Epidemiologic Wave

The first Houston Methodist patient infected with an Omicron variant was identified at the end of November 2021, a time when the Delta VOC was responsible for all COVID-19 cases in metropolitan Houston[6]. During this period, the metropolitan area was experiencing a steady decrease in total number of new COVID-19 cases (**Figure 1, Figure 2**).

Omicron increased in frequency unusually rapidly over a three-week period in December (**Figure 1, Figure 2).** By December 18, the genome sequence data showed that Omicron accounted for 90% of all new COVID-19 cases in our healthcare system (**Figure 2**). This represents the fifth wave of COVID-19 cases in metropolitan Houston (**Figure 1**). The estimated case doubling time during this three-week period was approximately 2.2 days (**Figure 2**), which means that Omicron increased in frequency approximately three times faster than Delta had increased in our area[6], an unprecedented trajectory for SARS-COV-2 infections.

Consistent with extensive infections caused by Omicron in southern Africa and elsewhere (https://www.cdc.gov/coronavirus/2019-ncov/variants/variant-

classifications.html, last accessed December 28, 2021; https://www.gov.uk/government/collections/new-sars-cov-2-variant, last accessed December 28, 2021), several patients had very recent travel histories to countries with a high prevalence of this VOC, suggesting acquisition of virus by some cases from abroad and importation into Houston. However, the great majority of Omicron patients had no documented travel outside the US and undoubtedly acquired the infection domestically, either in Houston or elsewhere.

To understand the geospatial distribution of Omicron in metropolitan Houston, patient metadata were acquired from the electronic medical record by standard informatics methods, and home address zip codes were used to visualize virus spread (**Figure 2**). The 862 Houston Methodist patients infected with Omicron during this period were distributed widely throughout metropolitan Houston, with 176 different zip codes represented (**Figure 2**). The widespread distribution of Omicron in the Houston metroplex in an extremely short period of time reflects the ability of this variant to spread unusually rapidly and effectively between individuals, and cause symptomatic disease.

Comparison of Omicron, Alpha, and Delta COVID-19 Cases

There is a considerable lack of detailed information about patients in the United States and elsewhere with COVID-19 caused by the Omicron VOC. We compared available metadata for all Houston Methodist patients infected with Omicron, Alpha, and Delta VOCs (**Table 1, Table 2**). The populations differed significantly in many characteristics,

including median age, hospital admission rates, maximum respiratory support, rate of vaccine breakthrough, and median length of stay (**Table 1, Table 2**).

Patients infected with Omicron were significantly younger than Alpha and Delta patients (**Table 1, Table 2**). Importantly, Omicron patients were hospitalized significantly less frequently than patients infected with either the Alpha or Delta variants, and had a significantly shorter median hospital length of stay (**Table 1, Table 2**).

We next analyzed Omicron vaccine breakthrough cases (**Table 1, Table 2**). We found 430 of the 862 total Omicron patients (49.9%) for whom we have whole genome sequence data met the CDC definition of vaccine breakthrough cases (**Table 1, Table 2**). There was no simple relationship between the time elapsed since administration of the second vaccination dose and the date of vaccination breakthrough. These 430 patients received either two doses of the Pfizer-BioNTech BNT162b2 (*n* = 299, 69%) or Moderna mRNA-1273 (*n* = 111, 26%), or one dose of J&J/Janssen JNJ-78436735 (*n* = 20, 5%) vaccine. This distribution reflects the majority use of BNT162b2 vaccination doses in our health system. Compared to either Alpha or Delta patients, a significantly greater percentage of patients with breakthrough cases was caused by the Omicron VOC (49.9% compared to 3.2% and 24.1% for Alpha and Delta VOCs, respectively) (**Table 1, Table 2**). We next analyzed individuals with breakthrough cases after receiving a third (booster) dose of either the Pfizer-BioNTech BNT162b2 or Moderna mRNA-1273 vaccine. We found that 85 (9.9%) of the 862 Omicron patients met this criteria. Consistent with Omicron causing a significantly increased number of vaccine breakthrough cases, it has been reported that this variant has reduced sensitivity to

antibody neutralization *in vitro*, likely in large part due to the extensive number of amino acid and other structural changes occurring in Omicron spike protein[10-29].

An Omicron Sublineage With Spike Protein Polymorphism L452R

The L452R amino acid change in spike protein of SARS-CoV-2 has arisen by convergent evolution in multiple genetic lineages throughout the course of the pandemic, most notably in the Delta VOC. L452R is located in the receptor binding domain of spike protein and has been associated with decreased antibody neutralization, escape from cell mediated immunity, and increased affinity for the ACE2 receptor[30, 31]. L452R was present in samples of Omicron collected in South Africa as early as November 11, 2021. We found that all 43 specimens collected over the first nine days after we identified Omicron in Houston Methodist patients had the 452R amino acid replacement. Our first L452 isolate was collected on day 10. By day 16, over half the Omicron isolates collected were L452, and the proportion of 452R isolates continued to diminish. Among the 65,109 Omicron samples deposited in GISAID as of 28 December 2021, only 727 (1.1%) have the 452R amino acid replacement and over half of these were from Houston Methodist patients. The initial high prevalence of Omicron containing 452R in spike protein in our population likely represents a founder effect of the original genotype introduced into Houston.

Spike-Gene Target-Failure Assay

To estimate Omicron variant frequency in patient samples not yet sequenced, we performed the TaqPath COVID-19 Combo Kit assay (ThermoFisher) on 657 samples collected from symptomatic patients between December 18, 2021 and December 22, 2021. In total, 604 (91.9%) of patient samples yielded a RT-PCR result with S-gene target-failure indicative of the Omicron variant. These data are consistent with the increasing frequency of new cases of COVID-19 caused by Omicron in our population (**Figure 2**).

Lack of Omicron Variant BA.2 in Houston Methodist Samples

The BA.2 Omicron sublineage was first identified in November 2021 in Australia in a patient who had traveled to South Africa (https://github.com/cov-lineages/pango-designation/issues/359; last accessed December 30, 2021). This sublineage does not have the full set of polymorphisms characteristic of BA.1 (B.1.1.529) and also has additional mutations unique to it (https://github.com/cov-lineages/pango-designation/issues/361; last accessed December 30, 2021). One important difference is that sublineage BA.2 lacks the spike protein deletion at amino acid 69/70 which means that it will not be detected by the SGTF assay. We inspected all full genome sequences present in our large database and this sublineage was not present. This includes all specimens taken from symptomatic patients or asymptomatic individuals. Thus, it is likely that routine use of an SGTF assay in our population will identify patients infected with Omicron.

Discussion

In this work, we describe information relevant to the surging Omicron wave in metropolitan Houston. In three weeks, Omicron was first identified in our population and rapidly increased to cause 90% of all new COVID-19 cases, with an estimated case doubling time of 2.2 days. The study was based on genome sequence analysis of 862 Omicron samples taken from socioeconomically, geographically, and ethnically diverse symptomatic patients. Several key findings were made, including (i) the Omicron VOC rapidly increased as a cause of COVID-19 and spread throughout the metroplex in an unusually short period of time, far faster than any other SARS-CoV-2 variant; (ii) Omicron caused significantly more vaccine breakthrough cases than the Alpha or Delta VOCs; (iii) Omicron patients were significantly younger than Alpha or Delta patients; (iv) significantly fewer Omicron patients required hospitalization compared to Alpha and Delta patients; (v) the median length of stay for hospitalized Omicron patients was significantly shorter than for Alpha and Delta patients, and consistent with this observation, the maximum respiratory support required for Omicron patients was significantly less than for Alpha or Delta patients. Our findings are largely consistent with many aspects of Omicron data reported from the UK, South Africa, and Canada[7-9, 32, 33] and are consistent with animal infection model data suggesting that Omicron causes less severe disease in mice and hamsters[34, 35]. This study was facilitated by a comprehensive and integrated population genomics and epidemiology project[2-6] implemented at the end of February, 2020, when the initial COVID-19 case was diagnosed in the Houston Methodist healthcare system.

Because we sequence the genome of approximately 90% of SARS-CoV-2 causing COVID-19 in our diverse Houston Methodist patient population, and have done so for almost two years, this means we are continuously monitoring the composition of this virus in a major US metroplex. This affords us the opportunity to rapidly assess changes in SARS-CoV-2 population genomic structure in the fourth largest city in the US. However, our study has several limitations. Although we sequenced the genomes of SARS-CoV-2 causing 90% of all Houston Methodist COVID-19 cases in the study period, this sample represents only approximately 5% of cases reported in the metropolitan region. Our patient population will underrepresent some demographic groups, for example homeless individuals and pediatric patients. The samples sequenced in this study were obtained from symptomatic individuals, which means that it is possible that we failed to identify Omicron subvariants or features preferentially represented in asymptomatic individuals.

In the aggregate, our data add critical new information to features of Omicron genomic epidemiology and patient characteristics in the US. Further, the present study highlights the importance of analyzing SARS-CoV-2 genome data integrated with patient metadata and stresses the need to continue to do this in near-real time as the Omicron surge continues, the virus evolves, and new variants with potentially altered fitness and biomedically relevant phenotypes are generated. Analyses of this type are also important in the context of vaccine formulation and long COVID, an increasing health and economic problem globally. Finally, the strategy we have used in this and previous studies[2-6] are readily applicable to future infectious diseases problems that warrant special attention.

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We declare that we have no conflict of interest.

Author Contributions

P.A.C., R.J.O., S.W.L., and J.M.M. had full access to all study data and take responsibility for the integrity of the data and the accuracy of the data analysis; concept and design by J.M.M., P.A.C., R.J.O., and S.W.L; data acquisition, analysis, or interpretation by all authors; drafting of the manuscript by all authors; statistical analysis by P.A.C.; funding obtained by J.M.M. and J.J.D.; and overall supervision by J.M.M. P.A.C., R.J.O., and S.W.L. contributed equally and are co-first authors.

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Figure Legends

Figure 1 Epidemiologic curve showing five COVID-19 disease waves in Houston Methodist patients. Number of new COVID-19 cases (y-axis) totals are shown as a +/- three-day moving average. Each of the five waves is shown in a different color. The first and second waves were composed of a heterogenous array of SARS-CoV-2 genotypes. The Alpha VOC shown in the third wave, the Delta VOC shown in the fourth, and the Omicron VOC shown in the fifth wave indicate their numeric prominence in those waves. The data should not be interpreted to mean that all cases in the third, fourth, and fifth waves were caused by Alpha, Delta, and Omicron VOCs, respectively. Rather, they are the dominant VOCs causing disease in Houston Methodist system patients in those waves. The fifth wave shown includes data through December 18, 2021. The figure was generated with Tableau version 2021.2.7 (Tableau Software, LLC, Seattle, WA), and is a modified version of one presented recently[6]. The curve is essentially superimposable on COVID-19 activity in all metropolitan Houston, Texas.

Figure 2 Increase in Omicron frequency over time and distribution in metropolitan Houston. The study time frame was November 27, 2021 through December 18, 2021. **A:** Omicron logistic growth model. The estimated case doubling time is 2.2 days. **B:** Cumulative increase in Omicron during the study period; y-axis is the cumulative number of new COVID-19 Omicron cases. At the end of the study period, Omicron caused 90% of all COVID-19 cases. **C – E:** Geospatial distribution of Omicron based on home address zip code for each patient. **B**: November 27 – December 3; **C**: November

27 – December 10; **D**: November 27 – December 18. Note differences in heat map scale for each panel. Figures were generated using Tableau version 2021.2.7. (Tableau Software, LLC, Seattle, WA).

572 **Table 1. Summary of pertinent patient metadata for 4021 unique patients infected**

573 **with Omicron or Alpha variants.**

	Omicron Variant	Alpha Variant	Total	Statistical Analysis
No. (%) with data	862 (21.4%)	3159 (78.6%)	4021	
Patient Characteristics				
Median Age (Years)	38.9	50.0	47.6	*P*<0.0001 Mann-Whitney
Female	505 (58.6%)	1623 (51.4%)	2128 (52.9%)	*P*=0.0002 Fisher's exact test
Male	357 (41.4%)	1536 (48.6%)	1893 (47.1%)	
Ethnicity				
Caucasian	256 (29.7%)	1242 (39.3%)	1498 (37.3%)	*P*<0.0001 Chi-square
Hispanic or Latino	156 (18.1%)	945 (29.9%)	1101 (27.4%)	
Black	368 (42.7%)	732 (23.2%)	1100 (27.4%)	
Asian	34 (3.9%)	123 (3.9%)	157 (3.9%)	
Other	3 (0.3%)	32 (1.0%)	35 (0.9%)	
Unavailable	45 (5.2%)	85 (2.7%)	130 (3.2%)	
BMI				
Median BMI	28.7	30.5	30.2	*P*<0.0001 Mann-Whitney
Admission Data				
Admitted	134 (15.5%)	1721 (54.5%)	1855 (46.1%)	*P*<0.0001 Fisher's exact test Odds Ratio: 0.154 (95% CI 0.126-0.187)
Not Admitted	728 (84.5%)	1438 (45.5%)	2166 (53.9%)	
Median LOS (Days) (Discharged patients only)	2.8	5.1	5.0	*P*<0.0001 Mann-Whitney
Max Respiratory Support				
ECMO	1 (0.7%)	7 (0.4%)	8 (0.4%)	*P*<0.0001 Chi-square
Mechanical Ventilation	6 (4.5%)	144 (8.4%)	150 (8.1%)	

Non-Invasive Ventilation	9 (6.7%)	164 (9.5%)	173 (9.3%)	
High Flow Oxygen	12 (9.0%)	364 (21.2%)	376 (20.3%)	
Low Flow Oxygen	42 (31.3%)	722 (42.0%)	764 (41.2%)	
Room Air	64 (47.8%)	320 (18.6%)	384 (20.7%)	
Mortality				
Alive	854 (99.1%)	2989 (94.6%)	3843 (95.6%)	*P*<0.0001 Fisher's exact test Odds Ratio: 0.165 (95% CI 0.081-0.336)
Deceased	8 (0.9%)	170 (5.4%)	178 (4.4%)	
Median PCR Cycle Threshold				
Abbott Alinity	19.5 n=327	22.4 n=1051	n=1378	*P*<0.0001 Mann-Whitney
Hologic Panther	22.1 n=60	24.2 n=359	n=419	*P*=.0991 Mann-Whitney
Vaccine				
Not Fully Vaccinated	432 (50.1%)	3058 (96.8%)	3490 (86.8%)	*P*<0.0001 Fisher's exact test Odds Ratio: 30.137 (95% CI 23.731-38.273)
Yes Fully Vaccinated	430 (49.9%)	101 (3.2%)	531 (13.2%)	

574 BMI: body mass index; CI: confidence interval; ECMO: extracorporeal membrane oxygenation; LOS: length of stay

575 **Table 2. Summary of pertinent patient metadata for 16,501 unique patients**

576 **infected with Omicron or Delta variants.**

	Omicron Variant	Delta Variant	Total	Statistical Analysis
No. (%) with data	862 (5.2%)	15639 (94.8%)	16501	
Patient Characteristics				
Median Age (Years)	38.9	48.2	47.7	*P*<0.0001 Mann-Whitney
Female	505 (58.6%)	8083 (51.7%)	8588 (52.0%)	*P*=0.0001 Fisher's exact test
Male	357 (41.4%)	7556 (48.3%)	7913 (48.0%)	
Ethnicity				
Caucasian	256 (29.7%)	6853 (43.8%)	7109 (43.1%)	*P*<0.0001 Chi-square
Hispanic or Latino	156 (18.1%)	4150 (26.5%)	4306 (26.1%)	
Black	368 (42.7%)	3445 (22.0%)	3813 (23.1%)	
Asian	34 (3.9%)	526 (3.4%)	560 (3.4%)	
Other	3 (0.3%)	112 (0.7%)	115 (0.7%)	
Unavailable	45 (5.2%)	553 (3.5%)	598 (3.6%)	
BMI				
Median BMI	28.7	29.5	29.5	*P*=0.0011 Mann-Whitney
Admission Data				
Admitted	134 (15.5%)	6730 (43.0%)	6864 (41.6%)	*P*<0.0001 Fisher's exact test Odds Ratio: 0.244 (95% CI 0.202-0.294)
Not Admitted	728 (84.5%)	8909 (57.0%)	9637 (58.4%)	
Median LOS (Days) (Discharged patients only)	2.8	5.4	5.3	*P*<0.0001 Mann-Whitney
Max Respiratory Support				
ECMO	1 (0.7%)	19 (0.3%)	20 (0.3%)	*P*<0.0001 Chi-square
Mechanical Ventilation	6 (4.5%)	720 (10.7%)	726 (10.6%)	

Non-Invasive Ventilation	9 (6.7%)	638 (9.5%)	647 (9.4%)	
High Flow Oxygen	12 (9.0%)	1778 (26.4%)	1790 (26.1%)	
Low Flow Oxygen	42 (31.3%)	2274 (33.8%)	2316 (33.7%)	
Room Air	64 (47.8%)	1301 (19.3%)	1365 (19.9%)	
Mortality				
Alive	854 (99.1%)	14815 (94.7%)	15669 (95.0%)	*P*<0.0001 Fisher's exact test Odds Ratio: 0.168 (95% CI 0.084-0.339)
Deceased	8 (0.9%)	824 (5.3%)	832 (5.0%)	
Median PCR Cycle Threshold				
Abbott Alinity	19.5 n=327	21.4 n=5081	n=5408	*P*<0.0001 Mann-Whitney
Hologic Panther	22.1 n=60	22.7 n=1291	n=1351	*P*=.9524 Mann-Whitney
Vaccine				
No vaccine	403 (46.8%)	11374 (72.7%)	11777 (71.4%)	*P*<0.0001 Chi-square
>7 days past 1st Vaccine	29 (3.4%)	489 (3.1%)	518 (3.1%)	
>14 days past 2nd Vaccine	345 (40.0%)	3641 (23.3%)	3986 (24.2%)	
>14 days past 3rd Vaccine	85 (9.9%)	135 (0.9%)	220 (1.3%)	

577 BMI: body mass index; CI: confidence interval; ECMO: extracorporeal membrane oxygenation; LOS: length of stay

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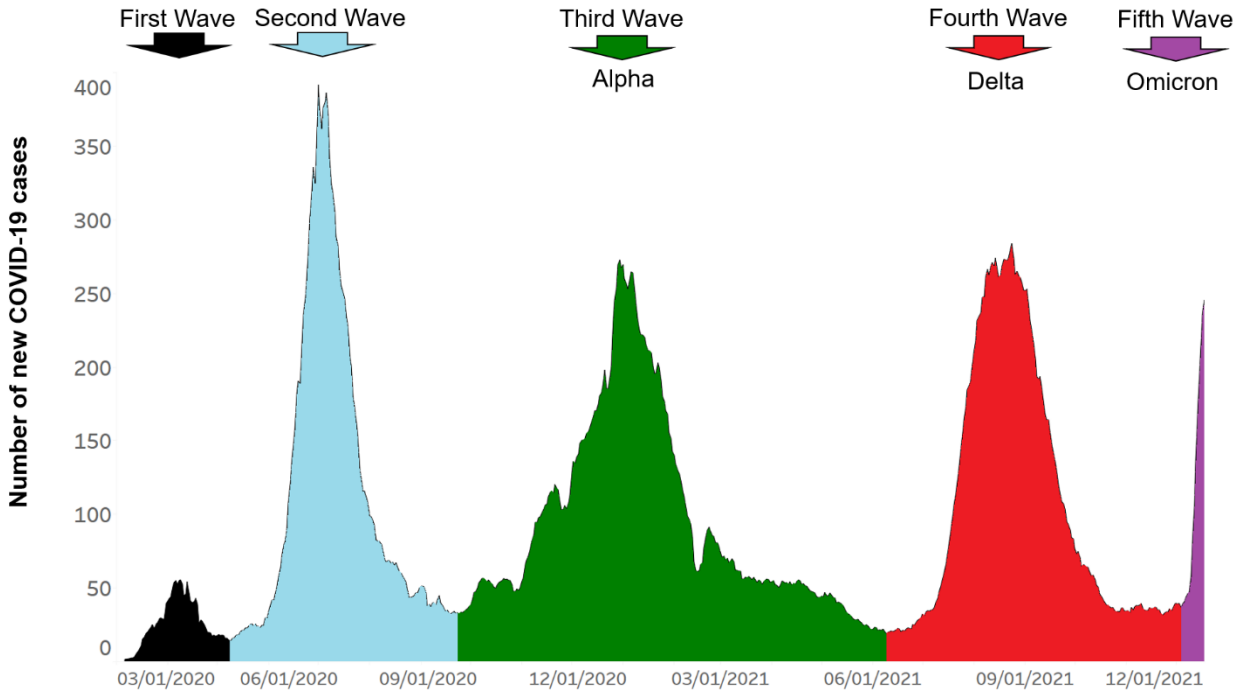
579 **Figure 1.**



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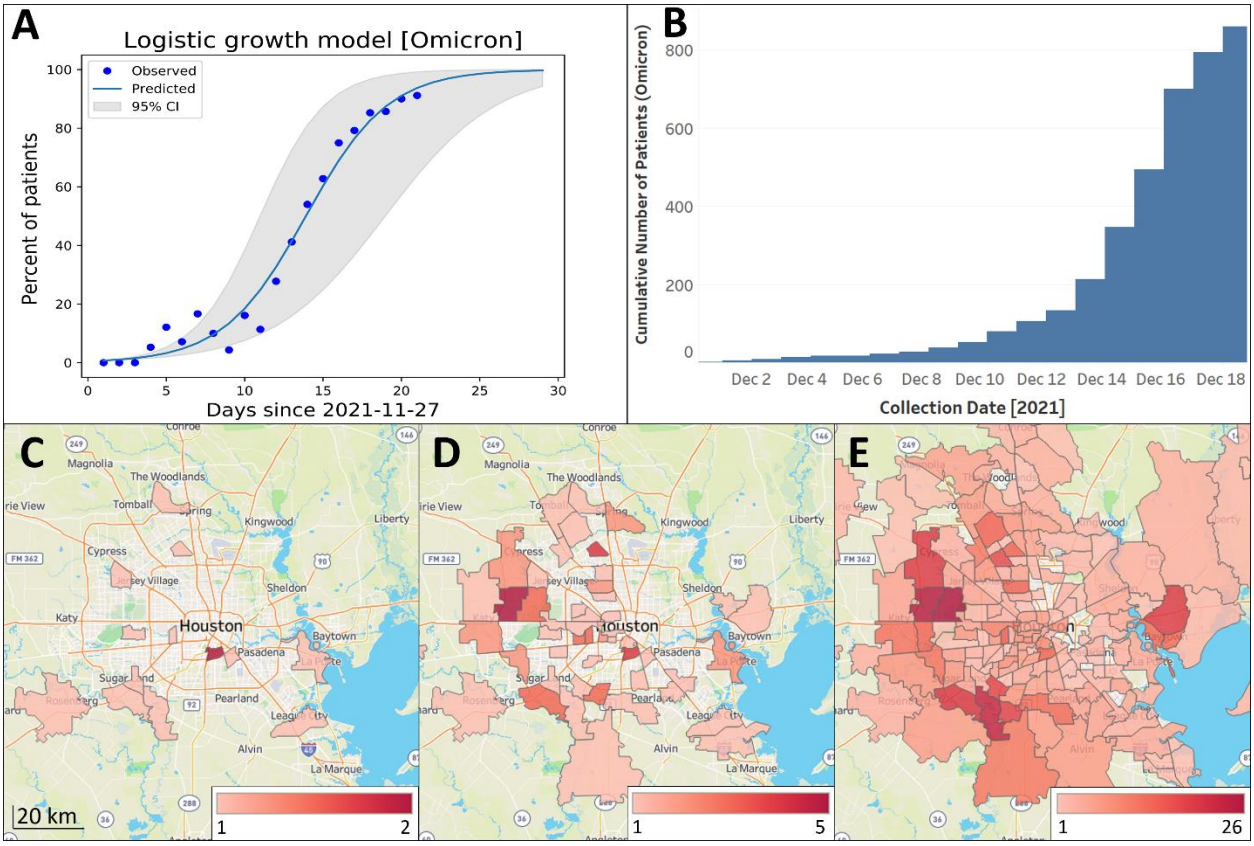
581 **Figure 2.**



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